

APPLICATION OF ANISE AND ROCKET ESSENTIAL OILS IN PRESERVATION OF OLD MANUSCRIPTS AGAINST FUNGAL DETERIORATION

Ahmed SAHAB^{1*}, Nagwa SIDKEY², Nermine ABED², Ayah MOUNIR³

¹ Department of Plant Pathology, National Research Center. Giza, Egypt.

² Microbiology Dept., Faculty of Science, Al-Azhar University, Cairo, Egypt.

³ Conservation and Microfilm Centre, General Book Organization of Egypt.

Abstract

The main goal of this work was to evaluate the effectiveness of zinc sulphate as a microelement in combination with anise and rocket essential oils as alternative preservatives protecting old manuscripts. Two fungal isolates of *Fusarium oxysporum* and *Trichoderma viride* isolated from deteriorated manuscripts were chosen to evaluate the protective agents. The obtained results revealed that the concentration of 100mM of zinc ion completely inhibit the growth of *F. oxysporum*, whereas the growth of *T. viride* was reduced growth by 80.22%. The essential oil of anise and rocket were chemically analyzed by GC-MS. The main components of anise were anethole (91.06%) followed by Cyclooctasiloxane (2.29%), then Humulen (1.48%). While, the main component of rocket identified were 1-Isothiocyanato-4-(methylthio) butane (erucin = sulforaphane) (81.23%), followed by Carvacrol (5.27%) and Thymol (5.16%). The results showed that the main mechanical properties either elongation percent (%) or maximum force (N/mm²) retained their values due to treatment with ZnSO₄ and fumigated by anise or rocket.

Keywords: Essential oils; Metallic ion; Manuscripts; *Fusarium oxysporum*; *Trichoderma viride*.

Introduction

Manuscripts constitute our most precious national and cultural heritage. Thus, the preservation of manuscripts is a serious problem for custodians throughout the world and this is why every possible effort must be taken to save these treasures for the future generation.

The responses of *Trichoderma* isolates to zinc ions were connected with concentrations of this metal. Zinc dosed 1000 and 3000ppm inhibited mycelial growth and *Trichoderma* spore germination, whereas lower concentrations of this metal stimulated *T. harzianum* growth [1], while Zn ion at 16ppm did not cause any effect on growth of *Trichoderma viride* [2], Zinc apparently inhibit the mycelial growth, whereas manganese ion stimulated spore germination of *T. viride* [3].

Anise (*Pimpinella anisum* L.), belonging to the *Umbelliferae* family is an annual herbaceous and a typical aromatic plant, which grows in several regions all over the world [4-6]. The chemical composition of essential oil of several *Pimpinella* species has been studied [7-11]. There are usually considerably variations in the major components within this species.

M. Embong et al. [7] established (E)-anethole (72.2%), (Z)-anethole (1.1%), anisyl ceton (0.9%), β-caryophyllene (0.8%) and carvone (0.3%); *M. Lawrence* [10] identified twenty-two

* Corresponding author: ahmedsahab2002@yahoo.co.uk

components of the anise oil with the major components of (E)-anethole (85%), (Z)-anethole (2.2%) and methyl chavicol (1.02%); *F. Askari et al.* [6] identified (E)-anethole (90%), eugenyl acetate (2%), γ -gurjunene (1.85%) and estragole (1.04%).

Volatile oils used for culinary, pharmaceutical and perfumery purposes are composed almost entirely of two classes of compounds, terpenes and phenylpropenes [12].

So, this investigation was done to evaluate effectiveness of zinc sulphate as a microelement and anis and rocket essential oils as alternative preservatives fumigant protecting old manuscripts.

Materials and Methods

Effect of zinc sulphate microelement on fungal growth and cellulolytic enzyme activity

This experiment was performed *in vitro* to study the effect of ZnSO₄ metallic ion on the growth and cellulolytic enzyme activity of *Fusarium oxysporum* and *Trichoderma viride* isolates. Sterile PDA solid medium was amended with serial concentrations of ZnSO₄ metallic ion, at concentrations of 10, 50, and 100mM. The tested microelement was added to the medium before autoclaving and a set of three replicates were used. The efficacy of the microelement was expressed as % of inhibition of mycelial growth against control, calculated by using formula by [13]. Three replicates were performed for each particular treatment.

$$IP = [(C - T)/C] \times 100 \quad (1)$$

where: IP - inhibitory percentage, C - Average colony diameter in check (control), T - Average colony diameter in treatment

Detection of cellulases activity

The previously selected fungal isolates were propagated on slants of Czapek's agar medium for seven days to prepare the spore suspension as inocula for the culture medium.

The activity of the cellulases was assayed according to *N. Sidkey et al* [14], 250mL Erlenmeyer flasks each contains 100mL buffer (citrate phosphate buffer, pH = 5.5), and 1% Avicel, CMC or filter paper, and 1.5% agar were boiled until it's ingredients completely homogenized. An aliquot of 20mL of the above homogenized substrate were poured in each petri dish. After solidification of the plates make 2 holes on each side of the agar plate and 0.1mL of the above cell free filtrate (CFF) were filled in the well. After incubation period of 20h at 28±2°C, the plates were flooded with iodine solution (1% iodine: 2% potassium iodide). The clear zone around the well was measured in mm, as the diameter of the zone increase as the cellulases activity increase.

Preparation of essential oil

Seeds (250g) of Anise (*Pimpinella anisum*), Fennel (*Foeniculum vulgare*), Rosemary (*Rosmarinus officinalis*), Rocket (*Eruca sativa*) and Tea tree oil (*Melaleuca alternifolia*) were obtained from Department of Medicinal and Aromatic Plants Research, National Research Center, Egypt were subjected to hydrodistillation for 3hr. using a Clevenger type apparatus to obtain essential oil according to the Egyptian Pharmacopoeia [15]. Lower aqueous layer was discarded and upper layer of oil was collected. The resulted essential oil of each treatment was separately dehydrated with anhydrous sodium sulfate and preserved in a sealed vial at 4°C until further analysis [16].

Analysis of essential oil

The GC-MS analysis of the essential oil samples was carried out using gas chromatography-mass spectrometry instrument stands at National Research Center with the following specifications, instrument: a TRACE GC Ultra Gas Chromatographs (THERMO Scientific Corp., USA), coupled with a THERMO mass spectrometer detector (ISQ Single

Quadrupole Mass Spectrometer). The GC-MS system was equipped with a TG-WAX MS column (30m x 0.25mm i.d., 0.25 μ m film thickness). Analyses were carried out using helium as carrier gas at a flow rate of 1.0 ml/min and a split ratio of 1:10 using the following temperature program: 40°C for 1 min; rising at 4.0°C/min to 160°C and held for 6min; rising at 6°C/min to 210°C and held for 1min. The injector and detector were held at 210°C. Diluted samples (1:10 hexane, v/v) of 0.2 μ L of the mixtures were always injected. Mass spectra were obtained by electron ionization (EI) at 70eV, using a spectral range of m/z 40-450.

Components were preliminary identified by comparison of their mass spectra with those of a computer library or with authentic compounds and confirmed by comparison of their retention indices, either with those of authentic compounds or with data published in the literature. The retention indices were calculated for all volatile constituents using a homologous series of alkanes [17].

Conservation treatments for fungal control

Preparation of paper sheets

Paper sheets made of cotton pulp were prepared according to method described in Figure1.

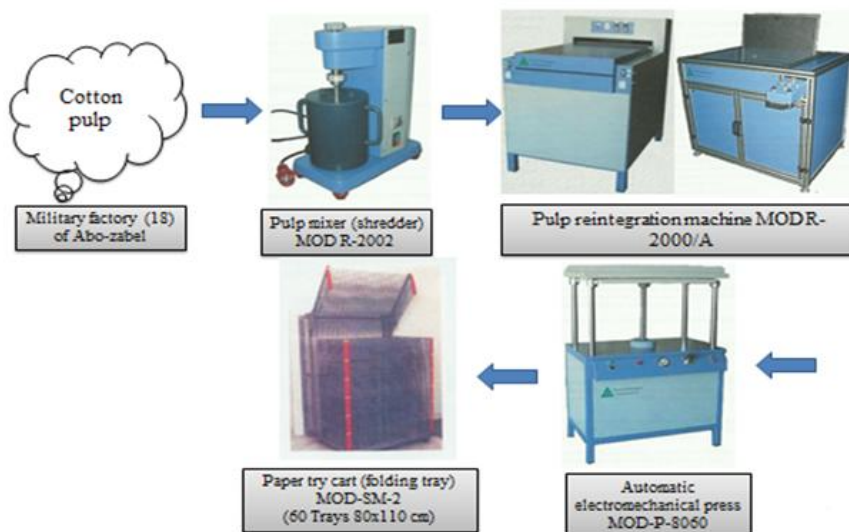


Fig. 1. Preparation of cotton paper to evaluate the effectiveness of zinc sulphate, anis and rocket essential oils

Treatment of paper sheet with protective materials (metallic ions and essential oils)

Eight paper sheets made of cotton pulp were cut into 15x20cm by using a scalpel, and sterilized by UV light exposure for one hour. Four sheets were soaked in a sterilized solution of ZnSO₄ at 0.05M concentration and left to dry under press. Spray 3 sheets with spore suspension of *Fusarium oxysporum* and other 3 sheets with spore suspension of *Trichoderma viride*. Incubate 2 sheets of paper for each fungus at a special jar containing anise essential oil and other 2 sheets with rocket essential oil as a fumigant, all sheets were incubated at 30°C, for 30 days. At the end of the incubation period, the following physical properties were estimated:

Tensile strength

Sample of infected cotton paper was standardized to (1.5x15cm) and tensile strength was tested using universal testing machine at National Institute for Standards (NIS), Egypt according to *M. Brindha et al* [18].

Scanning electron microscope (SEM)

This examination was performed to evaluate the distribution and infection of fungi on the surface of treated and non-treated cotton paper samples by using SEM Model Quanta 250 FEG (Field Emission Gun), at the Egyptian mineral resources authority, Central Laboratories Sector [19].

Statistical analysis

The collected data were statistically computed using the software Mstate-c for Windows. Results were expressed with the standard error of the treatment means for 95% confidence limits.

Results and Discussions

Effect of Zinc ion on growth and cellulolytic activity

This experiment was performed *in-vitro* to study the effect of ZnSO₄ metallic ion on the growth and cellulolytic enzyme activity of *Fusarium oxysporum* and *Trichoderma viride* isolates.

Data in Table 1 and Figure 2a and b showed that ZnSO₄ completely inhibited the *F. oxysporum* growth at 100mM concentration, but the inhibition was only up to 37.41 and 86.11% at 10 and 100mM concentration respectively. Whereas, ZnSO₄ amended solid medium at different concentrations of 10, 50 and 100mM of *T. viride* recorded percent inhibition to linear growth by 40.56, 57.22 and 80.22% respectively.

Also, all the cellulolytic activities including avicelase, carboxymethyl cellulases and filter paperase of *Fusarium oxysporum* as well as *Trichoderma viride* were adversely affected by the addition of ZnSO₄ to the medium. As, data of Avicelase enzyme showed that ZnSO₄ at 10, 50 and 100 mM added to the medium significantly decreased the enzyme activity of *F. oxysporum* by 12.47, 14.71 and 30.00% and enzyme activity of *T. viride* by 19.77, 24.42 and 27.33% respectively.

Table 1. Effect of Zinc ion on linear growth and cellulolytic activity of *Fusarium oxysporum* and *Trichoderma viride*

Metal ion	Conc. (mM)	linear growth (Ømm)		Cellulolytic activity {clear zone method (Ømm)}					
		<i>F. oxysporum</i>	<i>T. viride</i>	Avicel _{ase}		CMC _{ase}		Fp _{ase}	
				<i>F. oxysporum</i>	<i>T. viride</i>	<i>F. oxysporum</i>	<i>T. viride</i>	<i>F. oxysporum</i>	<i>T. viride</i>
Control		90.00	90.00	17.00	17.20	18.00	18.00	13.30	13.90
		A	A	A	A	A	A	B	A
10		56.33	53.50	14.88	13.80	14.30	13.30	0	10.00
		B	B	BC	BCD	B	BC	D	C
50		12.50	38.50	14.50	13.00	13.00	12.50	0	10.00
		E	C	BCD	CD	BC	C	D	C
100		0	17.80	12.90	12.50	13.00	11.80	0	10.00
		F	D	CD	D	BC	C	D	C

- Each figure represents average of three replicates, incubated at 30°C for 9days (solid) and 18days (liquid) Capek's medium.

- In each column, values followed by the same letters don't differ significantly (P≥0.05) (Duncan's multiple ranges test).

- Zero (0): No growth or no activity

Concerning, the effect of ZnSO₄ on the activity of CMC_{ase} of *F. oxysporum* as well as *T. viride* the same trends were also observed. However, the enzyme activity of Filter paperase, of *Fusarium oxysporum*, was completely inhibited by the addition of zinc sulphate at different

concentrations; on the other hand, it was found that, the addition of different concentrations of zinc ion led to the reduction of enzyme activity of *T. viride* by 28.06%. *G. Sanjeev and A. Eswaran* [20] and *G. Chand et al* [21] found the same trends, and agreement with other scientific researchers, of *G. Viniestra et al* [22] who stated that; the salts had been noticed directly related to metabolism, stimulation or inhibiting enzyme production in microorganism. Other scientists [23] reported that, cellulases enzymes of *T. reesei* were inhibited by Cu^{2+} (>20mM), Zn^{2+} (>1.0mM). However, metal cation of Zn^{2+} was necessary for cellulases synthesis by *T. viride* as reported by Mandels and Reese who hypothesized that, the metal may prevent some components necessary for induction from leaking out of the cells [24].

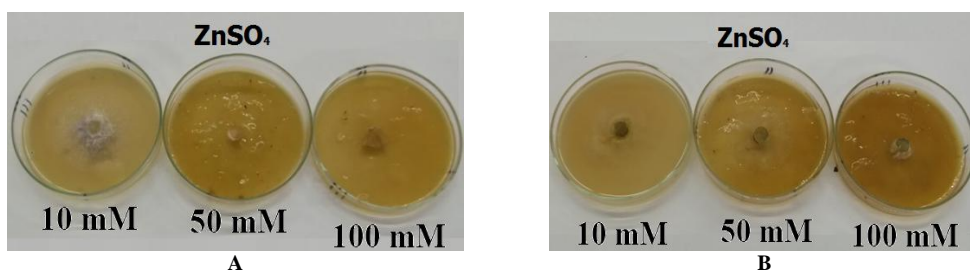


Fig. 2. Effect of Zinc sulphate ($ZnSO_4$) on linear growth (mm) of *Fusarium oxysporum* (A) & *Trichoderma viride* (B) isolates

Effect of some essential oils fumigation and $ZnSO_4$ treatments on paper quality

Trials were made to find out the most suitable procedure for protecting old manuscripts and documents from different deteriorations, which suffer through aging.

Identification and quantification of the constituent of anise

The chemical composition of anise (*Pimpinella anisum*) essential oil obtained by GC/MS analysis is presented in Table 2 and Figure 3, the main component identified were Anethole (91.06%) followed by Cyclooctasiloxane (2.29%), then Humulen (1.48%).

Table 2. GC-MS of *Pimpinella anisum* (anise)

No.	Retention time	Area%	Compound Name
1	4.80	0.09	α -Pinene
2	7.71	0.10	Benzeneacetic acid, α ,3,4tris[(trimethylsilyl)oxy], trimethylsilyl ester
3	8.71	0.03	α -Terpinen
4	10.34	0.25	Linalool
5	11.73	0.03	Geyrene
6	14.57	0.92	Benzene,1-methoxy-4-(2-propenyl)(P- Allylanisol)
7	16.85	0.30	Cyclohexasiloxane, dodecamethyl
8	18.28	91.06	Anethole
9	24.58	0.10	Aromandendrene
10	25.28	0.10	4,4-Dimethyl-3-(3-methylbut-3-enylidene)-2-methylenebicyclo[4.1.0] heptane
11	25.78	1.48	Humulen(v1)
12	26.64	0.06	α -Longipinene
13	27.39	0.08	Isolongifolene, 4,5dehydro
14	30.46	2.29	Cyclooctasiloxane, hexadecamethyl- CYCLOOCTA SILOXANE
15	36.11	0.71	Cyclononasiloxane, octadecamethyl
16	40.97	0.28	Octasiloxane,1,1,3,3,5,5,7,7,9,9,11,11,13,13,15,15 hexadecamethyl
17	49.42	0.15	Cyclonona siloxane, octadecamethyl
18	56.68	0.15	Heptasiloxane, hexadecamethyl

The same results were also reported before [5, 6, 16, 25-27]. Moreover, each of the essential oil components has its own contribution on biological activity of the oil. For example, Anethole was found in anise as the main compound, and this compound has more fungicidal effect [28].

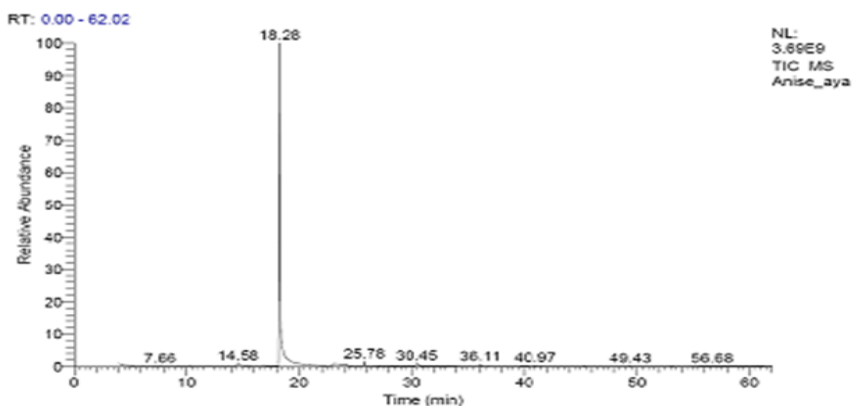


Fig. 3. GC/MS of *Pimpinella anisum* (anise) showed the highest peak (anethole)

Identification and quantification of the constituent of rocket essential oil

The chemical composition of rocket essential oil obtained by GC/MS analysis are presented in Table 3 and Figure 4, the main component identified were 1-Isothiocyanato-4-(methylthio) butane (erucin = sulforaphane) (81.23%), followed by Carvacrol (5.27%) and Thymol (5.16%). This result was also confirmed by [29-31].

Also, *B. Sabry* [32] reported that, erucin, which accounted for approximately 78.69% of the rocket extracts which play an important role as an antifungal agent

Table 3. GC/MS of *Eruca sativa* (rocket)

No.	Retention time	Area%	Compound Name
1	4.02	0.16	Allyl Isothiocyanate
2	6.28	0.94	9,10 Dideutero octadecanal
3	7.52	0.18	Cymene
4	7.74	0.23	Eucalyptol(1,8Cineole)
5	8.20	0.47	Undecane
6	8.39	0.19	Dodecane
7	9.89	0.29	l-Gala-l-ido-octose
8	10.42	0.16	Linalool
9	12.30	0.49	Camphor
10	12.83	0.18	Verbenol
11	13.28	0.24	Borneol
12	13.61	1.14	Terpinen-4-ol
13	15.08	1.94	B-D-Glucopyranose, 1thio-, 1-[N-hydroxy-5 (methylthio)pentanimidate]
14	16.54	0.85	Carvone
15	17.03	0.32	Eicosane
16	17.87	0.28	l-Gala-l-ido-octose
17	18. 41	5.16	Thymol
18	18.77	5.27	Carvacrol
19	24.45	81.23	Erucin (1-Isothiocyanato-4-(methylthio)butane)
20	25.87	0.32	β-D-Glucopyranose, 1-thio-1-[N-hydroxy-5-(methylthio) Pentanimidate] (Thiofanox)
21	27.72	0.29	1, 4, 7-Triazaheptane, 1,7-Bis(1-Methyl-1- phosphonato)Ethyl

Therefore, the effects of both essential oils and metallic ion on the susceptibility of recently paper to infection with fungi and on main mechanical properties were tested.

Results given in Table 4 and Figure 5 illustrated that, the infection of paper with *F. oxysporum* affected greatly the main mechanical properties both elongation percent (%) and maximum force (N/mm²) of the paper as they decreased from 0.675% and 11.5 to 1.644% and 23.16, respectively.

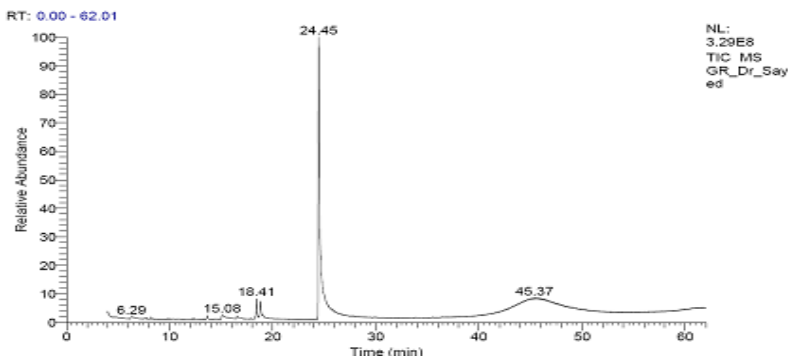


Fig. 4. GC/MS of *Eruca sativa* (rocket) showed the highest peak (1-Isothiocyanto-4 (methylthio) butane)

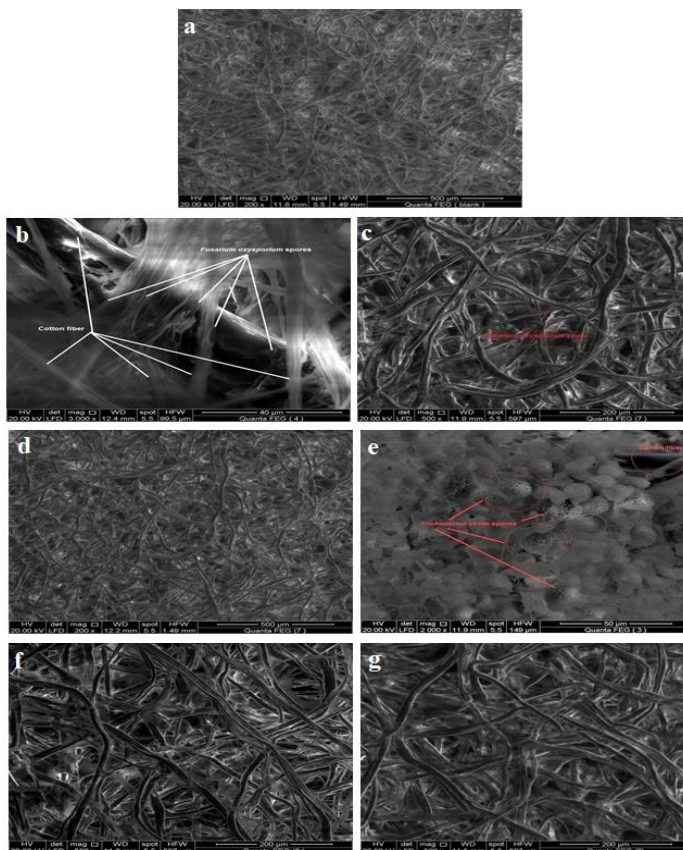


Fig. 5. SEM photographs showing the effect of ZnSO₄ and anise, rocket essential oil on cotton paper preservation: a - SEM picture at 200X. (control); b - SEM picture at 3000X. Cotton paper infected with *F. oxysporum* after 30days of infection. Arrows indicate the fungal spores (macro and micro conidia), c - SEM picture at 500X. Cotton paper infected with *F. oxysporum* and treated with ZnSO₄ and fumigated by anise. Arrows indicate fungal spore after 30 days of infection, d - SEM picture at 200X. Cotton paper infected with *T. viride* and treated with ZnSO₄ and fumigated by anise after 30 days of infection, e - SEM picture at 2000X. Cotton paper infected with *T. viride* after 30days of infection. Arrows indicate the fungal hypha and germinated spores, f - SEM picture at 500X. Cotton paper infected with *F. oxysporum* and treated with ZnSO₄ and fumigated by rocket after 30days of infection, g - SEM picture at 500X. Cotton paper infected with *T. viride* and treated with ZnSO₄ and fumigated by rocket after 30 days of infection.

Table 4. Effect of treatment with ZnSO₄ and fumigation by anise and rocket essential oils on the percentage of infection by either *Fusarium oxysporum* or *Trichoderma viride*, and subsequently on the physical properties of paper

Microorganism	<i>Fusarium oxysporum</i>		<i>Trichoderma viride</i>	
	Physical properties			
	Elongation%	Maximum force (N/mm ²)	Elongation%	Maximum force (N/mm ²)
Control	1.644	23.16	1.644	23.16
Infected Paper	0.675	11.52	0.985	12.59
Paper treated with ZnSO ₄ and fumigated by anise	2.182	25.42	1.164	13.62
Paper treated with ZnSO ₄ and fumigated by rocket	1.652	25.75	2.302	28.92

Moreover, the main mechanical properties, either elongation percent (%) or maximum force (N/mm²) retained their values due to treatment with ZnSO₄ and fumigated by anise or rocket. Since, infected paper with *F. oxysporum* treated with ZnSO₄ + anise, the elongation (%) and maximum force (N/mm²) attained 2.182% (223.3% increase) and 25.42 (120.65% increase) than the control, respectively. The same trend was also observed with rocket essential oil as the elongation (%) and maximum force (N/mm²) in the treated paper were increased by (144.7, 123.52 % increase), respectively.

Whereas, infected sheets with *T. viride* applied with ZnSO₄ and fumigated by anise or rocket essential oils revealed the same trend, as the elongation (%) was increased from 0.985% to 1.164% (18.2% increase) in anise and from 0.985% to 2.302% (133.7% increase) in rocket treatment, respectively. It is clear that almost all the maximum force and elongation values of the treated samples were higher than that of the infected paper with both fungi. The improvement in elongation and maximum force as a result of essential oils and metal ions may be due to the inhibition of fungi by essential oils and metal ions.

Scanning electron microscope (SEM)

The distribution and infection of *F. oxysporum* and *T. viride* on the surface of treated and non-treated cotton paper samples were observed in Figure 5.

Conclusion

It was revealed from the study that both fungicides and extracts of plant origin caused inhibition in mycelial growth and spore germination of *F. oxysporum* as well as *T. viride*.

So, it is worthy to state that, the recommendation to employ such treatments is not only due to their vital effectiveness in protecting treated paper sheets against biodegradation, but also to their advantage in elimination of harmful microorganisms which cause various health hazards in man and other organisms.

References

- [1] J. Dłużniewska, *Reaction of fungi of Trichoderma genus to selected abiotic factors*, **Electronic Journal of Polish Agricultural Universities (EJPAU)**, **6(2)**, 2003, p. #04.
- [2] M. Mandels, E.T. Reese, *Induction of cellulase in Trichoderma viride as influenced by carbon source and metals*, **Journal of Bacteriology**, **73(2)**, 1957, pp.269-278.
- [3] Z. Sierota, *Wpływ niektórych soli mineralnych na rozwój Trichoderma viride Pers. ex Fr. in vitro [Effect of some mineral salts on Trichoderma viride Pers. ex Fr. development in vitro]*, **Prace Instytut Badawczy Leśnictwa 611**, 1982, pp. 67-78 [In Polish].

- [4] R. Omidbaigi, A. Hadjiakhoondi, M. Saharkhiz, *Changes in content and chemical composition of Pimpinella anisum oil at various harvest time*, **Journal Essential Oil Bearing Plants**, **6**(1), 2003, pp. 46-50.
- [5] V.M. Rodrigues, P.T.V. Rosa, M.O.M. Marques, A.S. Petenale, M.A.A. Meireles, *Supercritical extraction of essential oil from aniseed (Pimpinella anisum L.) using CO₂ solubility, kinetics and composition data*, **Journal of Agricultural Food Chemistry**, **51**, 2003, pp. 1518-1523.
- [6] F. Askari, F. Sefidkon, V. Mozafarian, *Essential oil composition of Pimpinella aurea D.C. from Iran*, **Flavour and Fragrance Journal**, **20**, 2005, pp. 115-117.
- [7] M.B. Embong, D. Hadzie, S. Molnar, *Essential oil in Alberta anise oil (Pimpinella anisum L.)*, **Canadian Journal of Plant Science**, **59**, 1977, pp. 681-688.
- [8] M. Ashraf, M.A. Siddiqui, M.K. Bhatti, *Studies on the essential oils of the Pakistan species of the family Umbelliferae*, **Pakistan Journal of Scientific and Industrial Research**, **23**, 1980, pp. 211-212.
- [9] R. Ivanic, K. Savin, F.V. Robinson, *Essential oil from Pimpinella serbica fruits*, **Planta Medica**, **48**(1), pp.1983, pp. 60-61.
- [10] D.M. Lawrence, *Progres in essential oils [Anise oil, Calamus oil, Cardamom oils]*, **Perfumer and Flavorist**, **8**, 1984, pp. 63-65.
- [11] K.H.C. Bas'er, T. Özek, *Essential oil of Pimpinella aromatic Bieb. from Turkey*, **Journal Essential Oil Research**, **8**, 1996, pp. 463-464.
- [12] R.K.M. Hay, P.G. Waterman, **Volatile Oil Crops: Their Biology Biochemistry and Production**, Longman Scientific and Technical, New York, 1993.
- [13] M. Farzaneh, J. Hadian, S. Peighami, A. Sharifi, M. Ghorbanpoor, *Evaluation of Antifungal Activity of some plantn Essential oils against the grey mold of apple caused by Botrytis cinerea*, **Agricultural Research: Water, Soil, Plant Agriculture**, **7**(3), 2007, pp. 1-10.
- [14] N.M. Sidkey, S.M. Shash, M.S.C. Ammar, *Biodegradation of water hyacinth through certain hydrolytic enzymed secreted by different fungal isolates*, **Al-Azhar Bulletin of Science**, **8**(2), 1997, pp. 579-594.
- [15] * * *, **Egyptian Pharmacopoeia**, General Organization for Governmental Printing Office, Ministry of Health, Cairo, Egypt, 1984, pp. 31-33.
- [16] M.M. Ozcan, J.C. Chalchat, *Effect of collection time on chemical composition of the essential oil of Foeniculum vulgare subsp. piperitum growing wild in Turkey*, **European Food Research and Technology**, **224**, 2006, pp. 279-281.
- [17] N.W. Davies, *Gas chromatographic retention indices of monoterpenes and sesquiterpenes on methyl silicone and carbowax 20M phases*, **Journal of Chromatography**, **503**, 1990, pp. 1- 24.
- [18] M. Brindha, N. Kurunji Kumaran, K. Rajasigamani, *Evaluation of tensile strength and surface topography of orthodontic wires after infection control procedures: An in- vitro study*, **Journal of Pharmacy and Bioallied Sciences**, **6**(Suppl 1), 2014, pp. 44-48.
- [19] A. Helms Christine, J. Camillo Martiny, B. K. Hofman-Bang Ahring, M. Kilstrup, *Identification of bacterial cultures from archaeological wood using molecular biological techniques*, **International Biodeterioration and Biodegradation**, **53**, 2004, pp. 79 – 88.
- [20] K. K. Sanjeev, A. Eswaran, *Efficacy of Micro Nutrients on Banana fusarium Wilt. (Fusarium oxysporum f. sp. cubense) and its Synergistic Action with Trichoderma viride, Notulae Botanicae Horti Agrobotanici Cluj-Napoca*, **36** (1), 2008, pp.52-54.
- [21] G. Chand, U.S. Jaiswal, A.K. Maru, *Effect of micronutrients on panama wilt of banana (Fusarium oxysporum f. sp. cubense) and its synergistic action with Trichoderma viride*, **Conference Proceedings of International Conference on Innovative Approaches in Applied Sciences and Technologies (iCiAsT-2016)**, Scientific Educational Research

- Society, Meerut-250 004 India & Faculty of Science, Kasetsart University Bangkok 10900 Thailand February 01-05, 2016, pp. 49-50.
- [22] G. Viniegra, E. Favela, C. Aguilar, S. Romero, G. Diaz, C. Augur, *Advantages of fungal enzyme production in solid state over liquid fermentation systems*, **Biochemical Engineering Journal**, **13**(2-3), 2003, pp.157-167.
- [23] K.N.G. Thomas, J.G. Zeikus, Comparison of Extracellular Cellulase Activities of *Clostridium thermocellum* LQRI and *Trichoderma reesei* QM9414, **Applied and Environmental Microbiology**, **42**(2), 1981, pp. 231-240.
- [24] M. Mandels, E.T. Reese, *Fungal cellulases and the microbial decomposition of cellulosic fabric*, **Journal of Industrial Microbiology and Biotechnology**, **22**, 1999, pp. 225-240.
- [25] G. Singh, I.P.S. Kapoor, P. Singh, C.S. de Heluani, C.A.N. Catalan, *Chemical composition and antioxidant potential of essential oil and oleoresins from anise seeds (*Pimpinella anisum* L.)*, **International Journal of Essential Oil Therapeutics**, **2**, 2008, pp. 122-130.
- [26] M. Acimovic, V. Tesevic, M. Todosijevic, J. Djisalov, S. Oljaca, *Compositional characteristics of the essential oil of *Pimpinella anisum* and *Foeniculum vulgare* grown in Serbia*, **Botanica Serbica**, **39**(1), 2015, pp. 9-14.
- [27] S. E. Aly, B. A. Sabry, M. Saad, A. S. Hathout, *Assessment of antimycotoxigenic and antioxidant activity of star anise (*Illicium verum*) in vitro*, **Journal of the Saudi Society of Agricultural Sciences**, **15**(1), 2016, pp. 20–27.
- [28] S. Takayuki, S. Mami, M. Azizi, F. Yoshiharu, *Antifungal effects of volatile compounds from black zira (*Bunium persicum*) and other spices and herbs*, **Journal Chemistry Ecology**, **33**, 2007, pp. 2123- 2132.
- [29] M. Schluter, R. Gmelin, *Abnormalc Enzymatische Spaltung von 4-Methylthiobutyl-glucosinolat in Frischpflanzen von *Eruca sativa**, **Phytochemistry**, **11**, 1972, p. 3427.
- [30] S.F. Vaughn, M.A. Berhow, *Glucosinolate hydrolysis products from various plant sources: pH effects, isolation, and purification*, **Industrial Crops and Products**, **21**, 2005, pp. 193–202.
- [31] M. Khoobchandani, K. Ojeswi, N. Ganesh, M. Srivastava, S. Gabbanini, R. Matera, R. Iori, L. Valgimigli, *Antimicrobial properties and analytical profile of traditional *Eruca sativa* seed oil: Comparison with Various Aerial and Root Plant Extracts*, **Food Chemistry**, **120**, 2010, pp. 217–224.
- [32] B. A. Sabry, *Evaluation of some plant extracts as antioxidants and protectors against the harmful effects of aflatoxins on albino rat*, **Ph.D. Thesis**, Faculty of Science, Benha University, Benha, Egypt, 2011.
-

Received: May 19, 2017

Accepted: June 09, 2018